

Prevalence and odds of *Staphylococcus aureus* carriage in atopic dermatitis: a systematic review and meta-analysis

J.E.E. Totté,¹ W.T. van der Feltz,² M. Hennekam,^{1,*} A. van Belkum,^{3,4} E.J. van Zuuren⁵ and S.G.M.A. Pasmans¹

¹Department of Dermatology and ⁴Department of Medical Microbiology and Infectious Diseases, Erasmus MC University Medical Centre, Rotterdam, the Netherlands

²Molecular and Cellular Life Sciences, Utrecht University, Utrecht, the Netherlands

³bioMérieux, Microbiology, La Balme Les Grottes, France

⁵Department of Dermatology, Leiden University Medical Centre, Leiden, the Netherlands

Summary

Correspondence

J.E.E. Totté.

E-mail: j.totte@erasmusmc.nl

*Current affiliation: Youth Health Care Rijmond, Rijmond, the Netherlands

Accepted for publication

8 March 2016

Funding sources

The Department of Dermatology of the Erasmus MC University Medical Centre Rotterdam received an unrestricted grant from Microcos Human Health, the Netherlands.

Conflicts of interest

The Department of Dermatology of the Erasmus MC University Medical Centre Rotterdam received an unrestricted grant from Microcos Human Health, the Netherlands. A.v.B. is an employee of bioMérieux, a company developing and selling infectious disease diagnostics. W.v.d.F. is a former employee of Microcos Human Health.

DOI 10.1111/bjd.14566

Background *Staphylococcus aureus* is increasingly implicated as a possible causal factor in the pathogenesis of atopic dermatitis (AD). However, the reported prevalence rates of skin and nasal colonization in the literature vary widely.

Objectives This study evaluates the prevalence and odds of skin and nasal colonization with *S. aureus* in patients with AD.

Methods A systematic literature search was conducted. Odds ratios (ORs) for colonization in patients vs. controls and the prevalence of colonization in patients were pooled using the random-effects model.

Results Overall, 95 observational studies were included, of which 30 had a control group. The Newcastle–Ottawa Scale was used to assess study quality, with the majority of studies being of fair to poor quality. Patients with AD were more likely to be colonized with *S. aureus* than healthy controls [OR 19.74, 95% confidence interval (CI) 10.88–35.81]. Differences were smaller in nonlesional skin (OR 7.77, 95% CI 3.82–15.82) and in the nose (OR 4.50, 95% CI 3.00–6.75). The pooled prevalence of *S. aureus* colonization among patients was 70% for lesional skin, 39% for nonlesional skin and 62% for the nose. In lesional skin, meta-regression showed that the prevalence of colonization increased with disease severity. Study heterogeneity should be taken into consideration when interpreting the results.

Conclusions These results demonstrate the importance of colonization with *S. aureus* in AD. Further evaluation of the mechanisms by which *S. aureus* influences inflammation is required in addition to the development of targeted strategies to decrease skin and nasal *S. aureus* load.

What's already known about this topic?

- *Staphylococcus aureus* colonizes the skin of patients with atopic dermatitis.

What does this study add?

- For the first time, data on *S. aureus* colonization in atopic dermatitis are systematically summarized showing an increased risk of colonization of lesional skin, nonlesional skin and the nose in patients vs. healthy controls.

Increased colonization with *Staphylococcus aureus* in the skin of patients with atopic dermatitis (AD) was first described in the 1970s. Multiple studies confirmed this finding, reporting a

prevalence of skin colonization with *S. aureus* ranging from around 30% to nearly 100%.^{1–4} The underlying pathogenic mechanisms of *S. aureus* in relation to AD have still not been

fully elucidated. However, recent studies suggest a causal role in the complex pathogenesis of AD by showing that *S. aureus* colonization precedes (flares of) the disease.^{5–9} *S. aureus* can facilitate skin barrier defects and inflammation in AD using different mechanisms.^{4,10} Examples of this include the stimulation of mast-cell degranulation by staphylococcal delta toxin, the induction of keratinocyte apoptosis by alpha toxin, the stimulation of T cells by enterotoxins that act as superantigens and the modulation of inflammation by staphylococcal surface proteins, protein A and lipoteichoic acid.^{10–14}

As *S. aureus* contributes to both skin barrier defects and inflammation, a more proactive control of *S. aureus* in certain patients may help to reduce disease severity. However, the use of antibiotics can result in resistance of *S. aureus* and perturbation of healthy microbiota, which has been shown to have potentially deleterious health effects.^{15–18} At present, new targeted antimicrobial therapies (such as lysins) are being developed, which are directed against single bacteria (e.g. Staphfect SA.100 against *S. aureus*).^{19–22} Therefore it is important to identify patients with AD who can potentially benefit from antistaphylococcal treatment.

Defining the prevalence of *S. aureus* skin and mucosal colonization in (subgroups of) patients with AD might provide more insight into the importance of *S. aureus* as a contributor to the disease and its severity.

Current prevalence rates of *S. aureus* colonization reported in AD vary widely, mainly depending on the type of patients included, the sample size and the methods used to collect and detect *S. aureus* or its products. The swab and the scrub method are frequently used to collect microorganisms from the skin.²³ Swabs collect bacteria from the superficial layer of the skin, whereas a scrub technique allows collection of superficial skin cells and associated microbes.²⁴ The detection of *S. aureus* was predominantly based on culture-based methods. In recent years DNA sequencing methods have allowed for the determination of the complete microbial composition at species level and recently upcoming metagenomics techniques can be used for identification at strain level.²⁵

In this systematic review we aim to provide an overview and a pooled estimate of the prevalence and odds of colonization with *S. aureus* in patients with AD.

Materials and methods

Type of study

Both experimental and observational (original, human) studies were included; however, case reports were excluded. No restrictions were made relating to publication date and language.

Type of participants

Patients of all ages with a diagnosis of AD confirmed by a physician were included.

Type of outcome measures

The primary outcome was the proportion of patients with presence of *S. aureus* on the skin (lesional and nonlesional) or in the nose. Secondary outcomes were (i) the presence of *S. aureus* virulence factors on the skin; and (ii) the relation between AD severity and colonization with *S. aureus*.

In intervention studies, only the baseline measurement was included in this review. When studies reported multiple measurements over time taken from the same skin site (without treatment regimen), or when multiple locations were sampled at the same time point, the mean was included in the meta-analysis. Studies that reported solely on methicillin-resistant *S. aureus* were excluded.

Search strategy

The search was conducted in Embase (from 1947), Medline (from 1946), OvidSP (from 1946), Pubmed (from 1947), Web of Science (from 1945) and The Cochrane Central Register of Controlled Trials (CENTRAL) up to 16 September 2014 (Table S1; see Supporting Information). A cross-reference check was performed to identify further relevant studies.

Study selection and data extraction

The titles and abstracts were screened for relevance. Articles or abstracts were selected based on predefined inclusion and exclusion criteria (Appendix S1; see Supporting Information). Non-English articles were translated by an official translation service when considered relevant. The methodological quality of the articles was rated using an adapted version of the Newcastle–Ottawa Scale (NOS).^{26,27} Uncontrolled studies could reach a maximum score of 7 points and studies including a control group could reach a maximum score of 8 points. Using a scoring algorithm (Appendix S2; see Supporting Information), the controlled studies were classified as being of poor, fair or good methodological quality, based on their NOS scores for patient selection, comparability and outcome.²⁸ Study selection and quality assessment was conducted independently by two researchers (J.E.E.T. and W.T.v.d.F., J.E.E.T. and M.H. or W.T.v.d.F. and M.H.). Disagreements were resolved and consensus was reached. If identical populations were described in different publications within an overlapping time period, the study with the most extensive reporting of results was included.

Statistical analysis

A meta-analysis was performed using a random-effects model. A weighted prevalence of colonization with *S. aureus* on the skin and in the nose was calculated. In controlled studies the prevalence of colonization in patients and controls was compared, expressed as an odds ratio (OR) with a 95% confidence interval (CI). Heterogeneity was assessed using I^2 . In cases of substantial heterogeneity between studies ($I^2 > 50\%$) the

reasons for heterogeneity were explored using meta-regression (using the unrestricted maximum likelihood method and in cases where there were more than 10 available studies) for the variables NOS score, age and AD severity. For the meta-regression on severity, studies that used the Eczema Area Severity Index (EASI) score or the SCORing Atopic Dermatitis (SCORAD) score were selected. Cut-off values for mild, moderate and severe AD were used as previously described.^{29,30} Subgroup analysis was performed for variables that were significant in the meta-regression. Additional subgroup analysis was carried out for studies in which patients were not receiving antibiotic treatment. All statistical analyses were performed using Comprehensive Meta-Analysis Version 2.2 (Biostat, Englewood, NJ, U.S.A.). Publication bias was evaluated using funnel plots, Egger's regression and the trim-and-fill method.³¹ The present systematic review was conducted and reported in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses guidelines.³²

Results

Study characteristics

The search yielded a total of 4909 articles, of which 2990 articles remained after deduplication. We used article title and abstract to identify 350 studies (Fig. 1). After reading the full article texts, 95 studies met our inclusion criteria. All studies had an observational design and 30 studies compared patients

with AD with healthy controls. In 77% of the studies AD was diagnosed by clinical assessment (dermatologist or another specialized physician). The other studies did not clearly report who diagnosed the patients. The overall percentage of male patients was 52% and the mean age was 14 years (range 0.8–68.9) based on 58 studies. A total of 11 studies measured disease severity using EASI, with nine studies reporting a mean EASI score [17.7 (range 4.5–51.6)]. Twenty-two of the 40 studies that used SCORAD reported a mean score [48.2 (range 13.5–73.5)]. The remaining studies did not measure the disease severity, used other measuring methods or did not report mean EASI or SCORAD values. Overall, 54% of the studies were conducted in Europe, 27% in Asia and 13% in the U.S. Study characteristics including the methods used to collect and identify *S. aureus* are described in Table S2 (see Supporting Information).

Quality of the studies

We rated the quality of the 30 articles that included a control group as good (n = 4), fair (n = 4) and poor (n = 22). The quality of the 65 uncontrolled studies varied from 1 to 6 points out of 7 points on the NOS (Table S2 and Appendix S3; see Supporting Information) The main reason for downgrading the quality of controlled studies was incomparability of the patient and control groups. Uncontrolled studies were mainly downgraded owing to a limited description of the methods used for collection and identification of

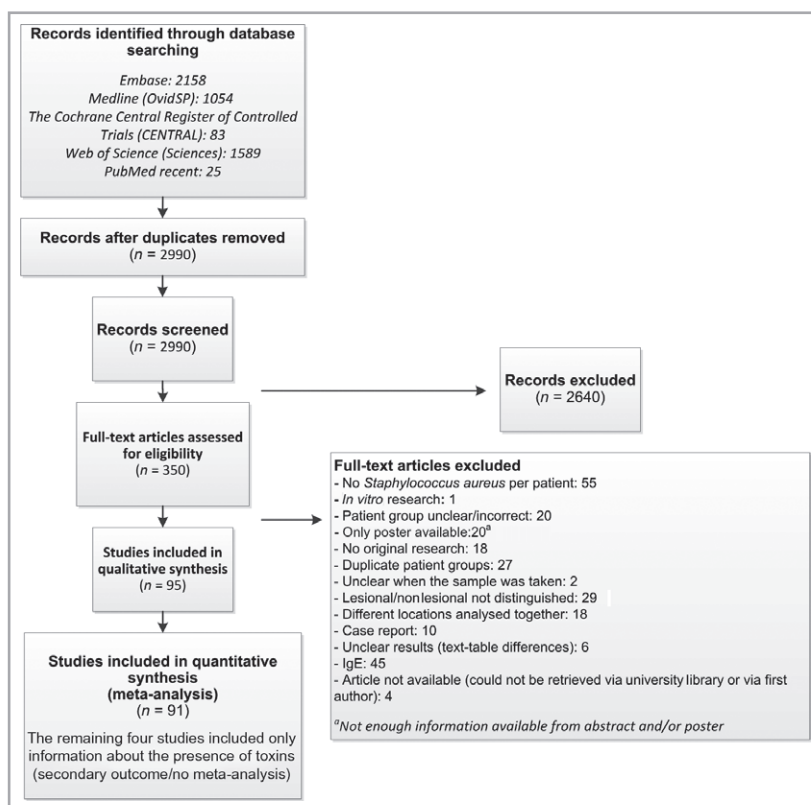


Fig 1. Flowchart of the search strategy and study selection.

S. aureus. The low NOS scores were also partly due to the inclusion of abstracts in this review, which provided limited information on methods.

Prevalence of nasal and skin colonization with *Staphylococcus aureus*

Overall, 81 studies (5231 patients) reported on colonization of the lesional skin and 30 studies (1496 patients) reported on colonization of the nonlesional skin. Pooled analysis showed that 70% of the patients with AD carried *S. aureus* on the lesional skin (95% CI 66–74; $I^2 = 88.31$) and 39% on the nonlesional skin (95% CI 31–47; $I^2 = 87.39$). Pooled results of 43 studies (2476 patients) that address nasal colonization estimated that 62% of the patients with AD carry *S. aureus* in the nose (95% CI 57–68; $I^2 = 85.20$) (Table 1 and Fig. S1; see Supporting Information). The prevalence varied substantially among studies (28–99% in lesional skin, 3–79% in nonlesional skin and 4–95% in the nose). This variation probably resulted in the considerable heterogeneity among studies and might be partly explained by variations in disease severity and the age of patients included in these studies.

Odds of colonization with *Staphylococcus aureus*

A total of 26 studies compared colonization of the lesional skin in patients with AD with healthy controls. From 10 studies the OR could not be obtained as the reported percentage of patients colonized with *S. aureus* or controls was either 100% or 0%. A pooled OR based on the remaining 16 studies (including 823 patients and 688 controls) showed that patients were significantly more likely than controls to be colonized with *S. aureus* on the lesional skin (OR 19.74, 95% CI 10.88–35.81; $P < 0.001$; $I^2 = 66.04$). Overall, 12 of 20 studies were eligible for inclusion in the pooled analysis for the nonlesional skin (550 patients and 446 controls) (OR 7.77, 95% CI 3.82–15.82; $P < 0.001$; $I^2 = 63.08$). Pooled analysis of 19 of 21 studies that evaluated nasal colonization (1051 patients and 1263 controls) showed that 57% of the patients were positive for *S. aureus* in the nose vs. 23% of the controls (OR 4.50, 95% CI 3.00–6.75; $P < 0.001$; $I^2 = 70.31$) (Table 2).

Meta-regression and subgroup analysis

Heterogeneity between the studies was considerable, mainly in the pooled analysis of prevalence (> 85%). A meta-regression for the variables AD severity, NOS score and age was performed to identify possible sources of heterogeneity. The prevalence of lesional skin colonization was independent of the NOS score but increased with AD severity (1.02, 95% CI 0.21–1.82) and age (0.64, 95% CI 0.15–1.14). A subgroup analysis of the studies that included patients with mild AD showed colonization of the skin in 43% of the patients (95% CI 31–57; $I^2 = 79.15$), whereas the pooled prevalence for severe AD was 83% (95% CI 74–89; $I^2 = 65.78$). For the

nonlesional skin, colonization decreased with a higher NOS score (–0.27; 95% CI –0.50 to –0.04). Subgroup analysis of the studies with a higher quality (NOS > 4) showed a pooled prevalence of 31% (95% CI 23–40; $I^2 = 64.62$), which is lower than the overall prevalence of 39%. Colonization of the nose was independent of the three variables (Table 1).

The ORs for colonization in patients with AD vs. controls were independent of the NOS and age. Severity was not tested as fewer than 10 studies that measured this variable were available (Table 2). Additional subgroup analysis, performed with studies that excluded patients who used antibiotics and corticosteroids at the time of inclusion, showed pooled ORs that were higher than the original pooled estimate of all studies (Tables 1 and 2).

Enterotoxins prevalence

The prevalence of at least one toxin-producing *S. aureus* strain on the lesional skin in patients varied between 31.5% and 80%. Staphylococcal enterotoxin B was the toxin found most often, with a prevalence of up to 70%. One study reported a prevalence of toxin-producing *S. aureus* of 11.5% in nonlesional skin. Three studies reported the presence of at least one toxin-producing *S. aureus* in the nose, with prevalence rates varying between 32% and 80%. Other studies reported combined results of skin and nose samples and were not taken into consideration in this study (Table S3 and Appendix S3; see Supporting Information).

Publication bias

The funnel plots for the prevalence of skin and nasal *S. aureus* in patients with AD showed asymmetry (Figure S2; see Supporting Information). The Eggers test confirmed the presence of publication bias with intercepts of 3.68 (95% CI 2.71–4.65, $P < 0.001$) for lesional skin, 0.76 (95% CI –3.06–4.85, $P = 0.69$) for the nonlesional skin and 2.63 (95% CI 0.84–4.42, $P = 0.005$) for the nose. Also the pooled analysis of the odds for colonization showed publication bias with an Eggers regression intercept of 2.47 (95% CI 1.66–3.28, $P < 0.001$) for lesional skin, 1.71 (95% CI 0.45–2.97, $P = 0.010$) for nonlesional skin and 2.08 (95% CI 0.64–3.52, $P = 0.023$) for the nose. Adjusted prevalence rates and ORs according to the trim-and-fill method were all lower than the original estimates (Table 1 and 2).

Discussion

In this systematic review we demonstrate that patients with AD are significantly more likely to be colonized with *S. aureus* than healthy controls on both the lesional and nonlesional skin and in the nose. Pooled prevalence of *S. aureus* carriage among patients is 70% for lesional skin, 39% for nonlesional skin and 62% for the nose. For lesional skin the prevalence appeared to be dependent on disease severity and age; however, this could

Table 1 Colonization with Staphylococcus aureus in patients with atopic dermatitis (AD)

	Number of studies	Pooled proportion of patients positive for colonization (95% CI)	Heterogeneity (I^2)	Pooled proportion of patients positive for colonization adjusted for publication bias	Meta-regression NOS, regression coefficient (95% CI)	Meta-regression Severe AD, regression coefficient (95% CI)	Meta-regression age, regression coefficient (95% CI)
Lesional skin							
All studies	81	0.70 (0.66–0.74)	88.31	0.57 (0.52–0.62)	0.07 (–0.10–0.24)	1.02 (0.21–1.82) ^{a,b}	0.64 (0.15–1.14) ^{b,c}
Studies including mild AD	4	0.43 (0.31–0.57)	79.15				
Studies including severe AD	9	0.83 (0.74–0.89)	65.78				
Studies excluding AB/steroid use	17	0.67 (0.58–0.75)	86.44				
Studies including age < 18 years	29	0.78 (0.71–0.84)	84.19				
Studies including age > 18 years	40	0.65 (0.59–0.71)	89.64				
Nonlesional skin							
All studies	30	0.39 (0.31–0.47)	87.39	0.38 (0.30–0.46)	–0.27 (–0.50 to –0.04) ^b	^d	0.76 (–0.01–1.52) ^e
Studies with a NOS score > 4	9	0.31 (0.23–0.40)	64.62				
Studies excluding AB/steroid use	11	0.24 (0.16–0.36)	85.22				
Nose							
All studies	43	0.62 (0.56–0.68)	85.20	0.53 (0.48–0.60)	–0.05 (–0.25–0.15)	0.62 (–0.15–1.39) ^f	0.12 (–0.47–0.72) ^g
Studies excluding AB/steroid use	8	0.58 (0.47–0.69)	78.23				

AB, antibiotics; CI, confidence interval; NOS, Newcastle–Ottawa Scale. All estimates were calculated using the random-effects model. Individual forest plots are provided in Figure S1 (see Supporting Information). ^aTwenty-eight studies included. ^bSignificant result. ^cSixty-nine studies included. ^dMeta-analysis was not performed because there were fewer than 10 studies. ^eFifteen studies included. ^fTwenty-seven studies included. ^gThirty-five studies included.

Table 2 Colonization with *Staphylococcus aureus* in patients with atopic dermatitis vs. healthy controls

	Number of studies	Pooled OR in patients vs. controls (95% CI)	Heterogeneity (I^2)	Pooled OR in patients vs. controls adjusted for publication bias	Meta-regression NOS, regression coefficient (95% CI)	Meta-regression Severity, regression coefficient (95% CI)	Meta-regression age, regression coefficient (95% CI)
Lesional skin							
All studies	16	19.74 (10.88–35.81) ^a	66.04	10.21 (5.44–19.16)	-0.05 (-0.47–0.37)		-0.55 (-1.84–0.74) ^b
Studies excluding AB/steroid use	6	27.43 (11.20–67.16) ^a	47.46				
Nonlesional skin							
All studies	12	7.77 (3.82–15.82) ^a	63.08	3.82 (2.18–6.72)			0.36 (-1.23–1.95) ^d
Studies excluding AB/steroid use	5	9.70 (3.60–26.13) ^a	51.06				
Nose							
All studies	19	4.50 (3.00–6.75) ^a	70.31		0.13 (-0.12–0.39)		0.68 (-0.48–1.84) ^f
Studies excluding AB/steroid use	7	5.54 (3.55–8.65) ^a	23.70				

AB, antibiotics; CI, confidence interval; NOS, Newcastle–Ottawa Scale. All estimates were calculated using the random-effects model. Studies that reported event rates of 0 or 1 were excluded as odds ratios (ORs) cannot be calculated with these event rates. Individual forest plots are provided in Figure S1 (see Supporting Information). ^aSignificant result. ^bFourteen studies included. ^cMeta-analysis was not performed because there were fewer than 10 studies. ^dTen studies included. ^eNo studies were trimmed according to the trim-and-fill method. ^fSixteen studies included.

not be confirmed for nonlesional skin or nasal colonization. Substantial to considerable heterogeneity, incomparability of patient and control groups, variation in methods used for sampling and poor description of exposures (such as treatment) downgraded the quality of the included articles, which should be taken into consideration when interpreting the results.³³

The typical features of AD skin, such as a compromised barrier integrity, altered sphingolipid metabolism and antimicrobial peptide expression probably facilitate colonization with *S. aureus*.^{34,35} The meta-regression analysis finds a higher prevalence of colonization among patients with more severe AD. However, the causal relationship between colonization with *S. aureus* and AD still has to be further clarified. Recent studies often suggest colonization with *S. aureus* as a primary cause rather than only a secondary effect of skin damage or an insufficient antistaphylococcal immune status. According to Kong et al. flares in AD accompany temporal microbial dysbiosis, dominated by *S. aureus*.⁵ Microbiome analysis of lesions in mice with an eczematous phenotype revealed that dysbiosis was a driving factor for dermatitis formation and bacterial inoculation experiments showed that *S. aureus* could accelerate eczematous inflammation.³⁶

Despite these studies that suggest a causal relationship, a systematic review by Bath-Hextall et al. did not demonstrate a beneficial clinical effect of untargeted anti-*S. aureus* therapy combined with steroids over steroids alone.^{37,38} However, other studies including treatment with mupirocin and bleach baths did show a reduction in clinical severity together with a reduction of *S. aureus* skin load.^{39,40} In our review we did not investigate the relationship between antistaphylococcal interventions and AD severity. We did conduct a subgroup analysis; including patients who were not receiving any antibiotic or corticosteroid treatment. This showed a lower prevalence of *S. aureus* on the skin and nose, which is not in line with the antibacterial effect of both antibiotics and corticosteroids.^{41,42} One explanation might be that the inclusion of patients who did not require treatment resulted in a selection of patients with mild AD who were less likely to be colonized with *S. aureus*.

Several natural and technical factors that are known to cause variation in microbiome outcomes might have influenced our results. There is variation between methods used to collect and detect *S. aureus* and its virulence factors on the human skin.^{43,44} Also, *S. aureus* might be present not only on the surface of the skin but also in deeper layers.⁴⁵ These differences highlight the importance of interpreting the results carefully, taking the methods used into consideration. Subgroup analysis for culture- vs. DNA-based detection methods were not performed owing to a small number of studies using DNA-based methods. Although DNA-based methods also include nonviable bacteria, they might provide more accurate results for quantifying *S. aureus* in the microbiome.

Furthermore, the impact of exposures such as treatment regimen and duration of the disease at the moment of collection were often poorly reported, which might have resulted in

performance bias. A subgroup analysis excluding patients who used antibiotics or steroids was performed to take the influence of treatment on the results into consideration. However, the use of other (aseptic) products might have also influenced the microbial composition. The duration of the disease might influence the activity of the host's immune response, which, in turn, could influence the presence of *S. aureus* via an antimicrobial effect.⁴⁶ The presence and quantity of microorganisms on the skin is influenced by many factors that naturally give rise to changes in the diversity of the microbiota over time and skin site (e.g. ethnicity and climate).^{24,47–52} It should be noted that our review reports on the proportion of *S. aureus* on the skin and mucosa determined at one specific time point.

As a result of underlying factors such as the (genetic) barrier defect and immune pathways enhancing a defective skin barrier, dysbiosis dominated by *S. aureus* is a chronic and recurring factor in AD.^{8,53–55} It is important to evaluate further the pathways by which *S. aureus* leads to inflammation and how current therapies already influence these pathways. Antibiotics and antiseptics are used in infected or severe AD.^{56–58} Functional textiles that are used as complementary treatment in AD might also decrease *S. aureus* colonization.⁵⁹ Glucocorticosteroids might also have an antibacterial effect besides their anti-inflammatory effect, probably via an effect on antimicrobial peptides, and even emollient monotherapy was shown to reduce bacterial colonization.^{9,60} The current use of antistaphylococcal therapies, together with literature that points to *S. aureus* as a driver in AD pathogenesis, underlines the importance of antistaphylococcal treatment in AD. However, long-term (preventive) use of antibiotics and glucocorticosteroids is undesirable as they can cause side-effects and antibiotic resistance.¹⁶

To date, this is the most comprehensive review that systematically summarizes data regarding *S. aureus* colonization in patients with AD. A large number of studies were included. These studies were mainly observational and often consisted of small numbers of patients. By not restricting the language of the search, selection bias was kept to a minimum. However, selection bias might have occurred owing to the exclusion of studies that did not report whether samples were taken from lesional or nonlesional skin. The covariate 'severity' in the meta-analysis was based on the level of the study, which may have led to an aggregation bias. As determining the prevalence of *S. aureus* colonization was not the primary objective in a substantial number of the studies, indirectness of evidence with regard to the study population might have occurred. Publication bias changed the outcomes considerably according to the trim-and-fill method. The quality of a large proportion of the individual studies was considered to be low. Future studies into the prevalence of *S. aureus* in patients vs. controls should take these quality criteria into consideration to raise the confidence in pooled estimates.

Despite the low quality of the studies included and the presence of publication bias, this systematic review and meta-analysis demonstrates that patients with AD are more frequently colonized with *S. aureus* than healthy controls and that

colonization is increased in more severe AD. These results provide an indication of the importance of colonization as a factor in the pathogenesis of AD and encourage evaluation of targeted antistaphylococcal therapy for the skin (and nose), for example based on the use of anti-*S. aureus* lysins. Prospective or experimental studies should further investigate causality and the mechanisms by which *S. aureus* colonization leads to inflammation. Host factors such as age and ethnicity, in addition to host–pathogen interaction, should be taken into consideration when investigating these mechanisms. The possible relevance of other microbes in the pathogenesis of AD should also be explored using metagenomic approaches. Additional examination of colonization in patients with different phenotypes (sensitized and nonsensitized, early onset vs. late onset) might provide insight into the type of patients who are likely to benefit most from targeted therapy against *S. aureus*.

Acknowledgments

We thank G.B. de Jonge (MSc), biomedical information specialist, for her assistance in the electronic literature search.

References

- 1 Park HY, Kim CR, Huh IS et al. *Staphylococcus aureus* colonization in acute and chronic skin lesions of patients with atopic dermatitis. *Ann Dermatol* 2013; **25**:410–16.
- 2 Matsui K, Nishikawa A, Suto H et al. Comparative study of *Staphylococcus aureus* isolated from lesional and non-lesional skin of atopic dermatitis patients. *Microbiol Immunol* 2000; **44**:945–7.
- 3 Leyden JJ, Marples RR, Kligman AM. *Staphylococcus aureus* in the lesions of atopic dermatitis. *Br J Dermatol* 1974; **90**:525–30.
- 4 Williams MR, Gallo RL. The role of the skin microbiome in atopic dermatitis. *Curr Allergy Asthma Rep* 2015; **15**:65.
- 5 Kong HH, Oh J, Deming C et al. Temporal shifts in the skin microbiome associated with disease flares and treatment in children with atopic dermatitis. *Genome Res* 2012; **22**:850–9.
- 6 Lebon A, Labout JA, Verbrugh HA et al. Role of *Staphylococcus aureus* nasal colonization in atopic dermatitis in infants: the Generation R Study. *Arch Pediatr Adolesc Med* 2009; **163**:745–9.
- 7 Bieber T. Atopic dermatitis. *Ann Dermatol* 2010; **22**:125–37.
- 8 Leung DY, Guttman-Yassky E. Deciphering the complexities of atopic dermatitis: shifting paradigms in treatment approaches. *J Allergy Clin Immunol* 2014; **134**:769–79.
- 9 Brussow H. Turning the inside out: the microbiology of atopic dermatitis. *Environ Microbiol* 2015; DOI: 10.1111/1462-2920.13050.
- 10 Travers JB. Toxic interaction between Th2 cytokines and *Staphylococcus aureus* in atopic dermatitis. *J Invest Dermatol* 2014; **134**:2069–71.
- 11 Nakamura Y, Oscherwitz J, Cease KB et al. *Staphylococcus aureus* δ -toxin induces allergic skin disease by activating mast cells. *Nature* 2013; **503**:397–401.
- 12 Brauweiler AM, Bin L, Kim BE et al. Filaggrin-dependent secretion of sphingomyelinase protects against staphylococcal α -toxin-induced keratinocyte death. *J Allergy Clin Immunol* 2013; **131**:421–7.e1–2.
- 13 Kotzin BL, Leung DY, Kappler J, Marrack P. Superantigens and their potential role in human disease. *Adv Immunol* 1993; **54**:99–166.
- 14 Gai C, Gonzalez C, Ledo C et al. Shedding of tumor necrosis factor receptor 1 induced by protein A decreases tumor necrosis factor

- alpha availability and inflammation during systemic *Staphylococcus aureus* infection. *Infect Immun* 2013; **81**:4200–7.
- 15 Chon SY, Doan HQ, Mays RM et al. Antibiotic overuse and resistance in dermatology. *Dermatol Ther* 2012; **25**:55–69.
 - 16 Chaptini C, Quinn S, Marshman G. Methicillin-resistant *Staphylococcus aureus* in children with atopic dermatitis from 1999 to 2014: a longitudinal study. *Australas J Dermatol* 2016; **57**:122–7.
 - 17 Antonov NK, Garzon MC, Morel KD et al. High prevalence of mupirocin resistance in *Staphylococcus aureus* isolates from a pediatric population. *Antimicrob Agents Chemother* 2015; **59**:3350–6.
 - 18 Lapin B, Piorkowski J, Ownby D et al. Relationship between prenatal antibiotic use and asthma in at-risk children. *Ann Allergy Asthma Immunol* 2015; **114**:203–7.
 - 19 Knoll BM, Mylonakis E. Antibacterial bioagents based on principles of bacteriophage biology: an overview. *Clin Infect Dis* 2014; **58**:528–34.
 - 20 Herpers BL, Badoux P, Pietersma F et al. Specific lysis of methicillin susceptible and resistant *Staphylococcus aureus* by the endolysin Staphfect SA.100™. Presented at the 24th European Congress of Clinical Microbiology and Infectious Diseases (ECCMID) 2014, Barcelona, Spain, 10–13 May 2014; abstr. R144.
 - 21 Herpers BL, Badoux P, Totté JEE et al. Specific lysis of *Staphylococcus aureus* by the bacteriophage endolysin Staphfect SA.100: in vitro studies and human case series. Presented at EuroSciCon: Antibiotic Alternatives for the New Millennium, London, U.K., 5–7 November 2014; 8.
 - 22 Totté JEE, van den Boogaard M, Pardo Cortes LM et al. *Staphylococcus aureus* specific topical treatment of rosacea and acne with the endolysin Staphfect SA.100. Presented at the 24th European Academy of Dermatology and Venereology Congress. Copenhagen, Denmark, 7–11 October 2015; abstr. P1212.
 - 23 Williamson P, Kligman AM. A new method for the quantitative investigation of cutaneous bacteria. *J Invest Dermatol* 1965; **45**:498–503.
 - 24 Grice EA, Kong HH, Renaud G et al. A diversity profile of the human skin microbiota. *Genome Res* 2008; **18**:1043–50.
 - 25 Garcia-Garcera M, Garcia-Etxebarria K, Coscolla M, et al. A new method for extracting skin microbes allows metagenomic analysis of whole-deep skin. *PLoS ONE* 2013; **8**:e74914.
 - 26 Wells G, Shea B, O'Connell D et al. The Newcastle–Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta-analyses. Available at: http://www.ohri.ca/programs/clinical_epidemiology/oxford.asp (last accessed 26 June 2016)
 - 27 Dowlatshahi EA, Wakke M, Arends LR, Nijsten T. The prevalence and odds of depressive symptoms and clinical depression in psoriasis patients: a systematic review and meta-analysis. *J Invest Dermatol* 2014; **134**:1542–51.
 - 28 McPheeters ML, Kripalani S, Peterson NB et al. Quality improvement interventions to address health disparities. Closing the quality gap: revisiting the state of the science. Available at: https://www.effectivehealthcare.ahrq.gov/ehc/products/386/1242/EvidReport208_CQGHHealthDisparities_FinalReport_20120824.pdf. (last accessed 26 June 2016).
 - 29 Leshem YA, Hajar T, Hanifin JM, Simpson EL. What the Eczema Area and Severity Index score tells us about the severity of atopic dermatitis: an interpretability study. *Br J Dermatol* 2015; **172**:1353–7.
 - 30 Oranje AP. Practical issues on interpretation of scoring atopic dermatitis: SCORAD Index, objective SCORAD, patient-oriented SCORAD and Three-Item Severity score. *Curr Probl Dermatol* 2011; **41**:149–55.
 - 31 Egger M, Davey Smith G, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ* 1997; **315**:629–34.
 - 32 Liberati A, Altman DG, Tetzlaff J et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate healthcare interventions: explanation and elaboration. *BMJ* 2009; **339**:b2700.
 - 33 Higgins JPT, Green S. *Cochrane Handbook for Systematic Reviews of Interventions* Version 5.1.0. Available at: <http://handbook.cochrane.org> (last accessed 26 June 2016).
 - 34 Ong PY, Ohtake T, Brandt C et al. Endogenous antimicrobial peptides and skin infections in atopic dermatitis. *N Engl J Med* 2002; **347**:1151–60.
 - 35 Arikawa J, Ishibashi M, Kawashima M et al. Decreased levels of sphingosine, a natural antimicrobial agent, may be associated with vulnerability of the stratum corneum from patients with atopic dermatitis to colonization by *Staphylococcus aureus*. *J Invest Dermatol* 2002; **119**:433–9.
 - 36 Kobayashi T, Glatz M, Horiuchi K et al. Dysbiosis and *Staphylococcus aureus* colonization drives inflammation in atopic dermatitis. *Immunity* 2015; **42**:756–66.
 - 37 Bath-Hextall FJ, Birnie AJ, Ravenscroft JC, Williams HC. Interventions to reduce *Staphylococcus aureus* in the management of atopic eczema: an updated Cochrane review. *Br J Dermatol* 2011; **164**:228.
 - 38 Bath-Hextall FJ, Birnie AJ, Ravenscroft JC, Williams HC. Interventions to reduce *Staphylococcus aureus* in the management of atopic eczema: an updated Cochrane review. *Br J Dermatol* 2010; **163**:12–26.
 - 39 Wong SM, Ng TG, Baba R. Efficacy and safety of sodium hypochlorite (bleach) baths in patients with moderate to severe atopic dermatitis in Malaysia. *J Dermatol* 2013; **40**:874–80.
 - 40 Lever R, Hadley K, Downey D, Mackie R. Staphylococcal colonization in atopic dermatitis and the effect of topical mupirocin therapy. *Br J Dermatol* 1988; **119**:189–98.
 - 41 Gong JQ, Lin L, Lin T et al. Skin colonization by *Staphylococcus aureus* in patients with eczema and atopic dermatitis and relevant combined topical therapy: a double-blind multicentre randomized controlled trial. *Br J Dermatol* 2006; **155**:680–7.
 - 42 Stalder JF, Fleury M, Sourisse M et al. Local steroid therapy and bacterial skin flora in atopic dermatitis. *Br J Dermatol* 1994; **131**:536–40.
 - 43 Fahlen A, Engstrand L, Baker BS et al. Comparison of bacterial microbiota in skin biopsies from normal and psoriatic skin. *Arch Dermatol Res* 2012; **304**:15–22.
 - 44 Gao Z, Tseng CH, Strober BE et al. Substantial alterations of the cutaneous bacterial biota in psoriatic lesions. *PLoS ONE* 2008; **3**:e2719.
 - 45 Nakatsuji T, Chiang HI, Jiang SB et al. The microbiome extends to subepidermal compartments of normal skin. *Nat Commun* 2013; **4**:1431.
 - 46 Chehoud C, Rafail S, Tyldsley AS et al. Complement modulates the cutaneous microbiome and inflammatory milieu. *Proc Natl Acad Sci USA* 2013; **110**:15061–6.
 - 47 Grice EA, Segre JA. The skin microbiome. *Nat Rev Microbiol* 2011; **9**:244–53.
 - 48 Lai Y, Cogen AL, Radek KA et al. Activation of TLR2 by a small molecule produced by *Staphylococcus epidermidis* increases antimicrobial defense against bacterial skin infections. *J Invest Dermatol* 2010; **130**:2211–21.
 - 49 Wanke I, Steffen H, Christ C et al. Skin commensals amplify the innate immune response to pathogens by activation of distinct signaling pathways. *J Invest Dermatol* 2011; **131**:382–90.
 - 50 Krishna S, Miller LS. Host–pathogen interactions between the skin and *Staphylococcus aureus*. *Curr Opin Microbiol* 2012; **15**:28–35.
 - 51 Kuehnert MJ, Kruszon-Moran D, Hill HA et al. Prevalence of *Staphylococcus aureus* nasal colonization in the United States, 2001–2002. *J Infect Dis* 2006; **193**:172–9.
 - 52 Sahoo KC, Sahoo S, Marrone G et al. Climatic factors and community – associated methicillin-resistant *Staphylococcus aureus* skin

- and soft-tissue infections – a time-series analysis study. *Int J Environ Res Public Health* 2014; **11**:8996–9007.
- 53 McAleer MA, Irvine AD. The multifunctional role of filaggrin in allergic skin disease. *J Allergy Clin Immunol* 2013; **131**:280–91.
- 54 Howell MD, Kim BE, Gao P *et al.* Cytokine modulation of atopic dermatitis filaggrin skin expression. *J Allergy Clin Immunol* 2009; **124** (3 Suppl. 2):R7–12.
- 55 Khattri S, Shemer A, Rozenblit M *et al.* Cyclosporine in patients with atopic dermatitis modulates activated inflammatory pathways and reverses epidermal pathology. *J Allergy Clin Immunol* 2014; **133**:1626–34.
- 56 Ring J, Alomar A, Bieber T *et al.* Guidelines for treatment of atopic eczema (atopic dermatitis) part I. *J Eur Acad Dermatol Venereol* 2012; **26**:1045–60.
- 57 Huang JT, Abrams M, Tloutan B *et al.* Treatment of *Staphylococcus aureus* colonization in atopic dermatitis decreases disease severity. *Pediatrics* 2009; **123**:e808–14.
- 58 Breuer K, Haussler S, Kapp A, Werfel T. *Staphylococcus aureus*: colonizing features and influence of an antibacterial treatment in adults with atopic dermatitis. *Br J Dermatol* 2002; **147**:55–61.
- 59 Lopes C, Silva D, Delgado L *et al.* Functional textiles for atopic dermatitis: a systematic review and meta-analysis. *Pediatr Allergy Immunol* 2013; **24**:603–13.
- 60 Angelova-Fischer I, Neufang G, Jung K *et al.* A randomized, investigator-blinded efficacy assessment study of stand-alone emollient use in mild to moderately severe atopic dermatitis flares. *J Eur Acad Dermatol Venereol* 2014; **28**:9–15.

Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher's website:

Appendix S1. Criteria for selecting studies.

Appendix S2. Quality assessment score.

Appendix S3. Supplementary references.

Fig S1. Meta-analysis. (a) Forest plot: odds of lesional skin colonization (all studies). (b) Forest plot: odds of nonlesional skin colonization (all studies). (c) Forest plot: odds of nasal colonization (all studies).

Fig S2. Publication bias. (a) Funnel plot of studies reporting prevalence of lesional skin colonization with *Staphylococcus aureus* in patients with atopic dermatitis (AD). (b) Funnel plot of studies reporting prevalence of nonlesional skin colonization with *S. aureus* in patients with AD. (c) Funnel plot of studies reporting prevalence of nasal skin colonization with *S. aureus* in patients with AD. (d) Funnel plot of studies reporting odds of lesional skin colonization with *S. aureus* in patients with AD. (e) Funnel plot of studies reporting odds of nonlesional skin colonization with *S. aureus* in patients with AD. (f) Funnel plot of studies reporting odds of nasal skin colonization with *S. aureus* in patients with AD.

Table S1. Digital search strategy (last updated 16 September 2014).

Table S2. Study characteristics including Newcastle–Ottawa Scale scores and results per study.

Table S3. Presence of enterotoxins in lesional skin of patients with atopic dermatitis and healthy controls.